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(64) Stabilized immunoglobulin and method of preparation.

(72) Histidine-stabilized immunoglobulin preparations and a method for their manufacture are disclosed. The present invention is particularly well suited for stabilization of human gamma globulin (IgG) preparations having a relatively low protein content. Preferred stabilized human gamma globulin preparations comprise about 5 wt-% or less gamma globulin, histidine at a concentration of about 0.025M to about 0.2M, and optionally glycine at a concentration of about 0.05M to about 0.5M. The pH value of the preparations is at least 6.0 but not more than 7.0. A pH value of about 6.4 is most preferred. Conductivity of the preparations is about 2 to about 4 millisiemens at 5°C., preferably about 2.5 to about 3.5 millisiemens at 5°C., and most preferably about 2.7 millisiemens at 5°C.

STABILIZED IMMUNOGLOBULIN
AND METHOD OF PREPARATION

Technical Field

5 This invention relates to stabilized immunoglobulin preparations. One aspect of this invention relates to stabilized highly purified immunoglobulin G preparations intended for injection into humans.

Background of the Invention

10 Under appropriate circumstances the injection of an antigen into an animal produces a specific antiserum that reacts selectively with the antigen. This antiserum contains proteins that are responsible for the recognition of the antigen, i.e.,
15 proteins that possess a so-called "antibody function." Such proteins are commonly referred to as "antibodies" or, in a broader sense, as "immunoglobulins" (Ig).

All individuals within a given species have
20 in common various Ig categories called "isotypes." For example, in humans 10 isotypes have been identified and grouped into five classes, namely IgG, IgM, IgA, IgD and IgE, which classes are further subdivided into subclasses, e.g., IgG₁, IgG₂,
25 IgG₃ and IgG₄ for Immunoglobulin G. These classes are also sometimes referred to or designated as γ G, γ M, γ A, γ D and γ E, respectively. The predominant serum immunoglobulins are of the class IgG or gamma globulins.

30 Gamma globulin (IgG) preparations intended for injection into humans originally were produced by the alcohol fractionation procedure developed by Dr. E. Cohn and coworkers of Harvard during the 1940's. This procedure is described in Cohn et al., J. Amer.
35 Chem. Soc. 68:459 (1946). These preparations were

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prepared as 10-18 wt-% protein solutions, the most common being a 15 wt-% protein solution. The latter was favored since at that concentration, the product exhibited the same viscosity as blood. Another advantage for these high protein gamma globulin products was their inherent shelf stability as judged by the absence of visually observable precipitation upon storage at 2-8°C. for time periods of 2-3 years. In contrast, preparations containing less than about 10 wt-% protein frequently exhibited visually observable precipitation in the final liquid product.

The foregoing, relatively high protein gamma globulin products initially were intended for intravenous (IV) administration. The latter is considered advantageous clinically because immediate high levels of circulatory protective antibodies are attainable, whereas approximately one-half of the antibody given intramuscularly (IM) is lost due to local proteolysis and incomplete absorption. Unfortunately, experience has shown the above product not to be safe for IV administration due to adverse anaphylatic reactions in recipients. °See, for example, Bowman, Clin. Obst. Gynec. 25:341 (1982) and Schroeder et al., Amer. J. Med., March 30, 1984, page 33. The formation of aggregated gamma globulin polymers during storage of products prepared by the standard cold alcohol procedure allows for these aggregates to combine with complement in the patient's blood and to produce an anticomplement reaction. Thus, all gamma globulin products produced by the standard cold alcohol procedure have been approved only for intramuscular use.

The formation of relatively large molecular weight aggregates of gamma globulin in aqueous

solutions is particularly deleterious to its pharmaceutical utility. Such denaturation is believed to be influenced by sulphhydryl-disulfide interchange reaction.

5 The ability of gammaglobulin to bind complement is greatly increased as a result of denaturation, in particular by aggregation to high molecular weight species. The complement binding mechanism of these aggregates appears to be identical to that of antigen-antibody complexes. Marcus, D.M., 10 (1960) J. Immunol. 84:273-284. In the case of IgG, it is known that the complement binding site requires two molecules close together. It is therefore possible that critical packing of the molecules is required, rather than any necessary conformational change. However, the size distribution of aggregates in denatured antibody solutions and its relation to anticomplement activity has not been examined.

 In earlier studies, Gerber, J. Immunol. 20 92:885-888 (1964) and Gerber, Arthritis Rheum. 17:85-91 (1971), it is postulated that in patients suffering from rheumatoid arthritis there is an in vivo mechanism triggered by metallic copper that causes increased formation of aggregate of gamma globulin. This worker found a beneficial effect to 25 the patient if the level of one amino acid, L-histidine, was increased. These studies were done at physiological pH values, i.e., a pH of about 7.4 however. Also, the effect of histidine protection on 30 relatively low protein human gamma globulin products in the area of absence of aggregate and anticomplementary activity was not examined in the studies by Gerber and cannot be predicted from the reported data.

35 As gamma globulin therapy has become an accepted practice, the demand for large quantities of

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commercial product worldwide has required manufacturers to adopt hyperimmunization programs for donors of the starting blood from which the gamma globulin-containing product is derived. The
5 resulting plasma or serum has a desired higher starting antibody titer. Hence, more product can be derived from a fixed starting volume of blood. While the desired final antibody titer can also be attained at a lower protein concentration than the previous
10 10-18 wt-% value, such gamma globulin-containing products are less stable during storage.

Thus, one must either add protein to the product to maintain the desired 10-18 wt-% concentration, or develop methods for stabilizing a
15 solution of the highly purified gamma globulin product having a protein concentration of less than 10 wt-%. For the former approach, it is common to add either purified albumin or gamma globulin derived from a low titer plasma source. However, this
20 approach is not economically attractive. Also, it exposes the patient to additional health risks since viral contaminants from the additional purified proteins are a possibility. An example of the latter approach can be found in U.S. Patent No. 4,186,196 to
25 Lundblad et al. where a relatively high concentration of maltose is added to the final buffer to stabilize a 5 wt-% gamma globulin product. The latter is derived from a modified cold alcohol procedure that is said to remove aggregates as well, the objective
30 being a product that can be administered intravenously.

Other prior attempts at stabilization of protein solutions are illustrated by U.S. Patent No. 2,826,533 to Fowell which discloses the use of
35 dextrose to increase the solubility of fibrinogen in

5 solution, U.S. Patent No. 4,089,949 to Thomas which discloses the use of a variety of carbohydrates (e.g., dextrose, mannose, galactose, fructose, lactose, sucrose and maltose) to enhance the solubility of an anti-hemophilic factor (AHF)-fibrinogen composition, and U.S. Patent No. 3,057,781 to Mace et al. which discloses stabilization of plasma with invert sugar.

10 Other workers have explored the possibility of producing a gamma globulin product by an entirely different process, again with an objective to produce a product suitable for IM or IV administration. The approach currently favored employs ion-exchange chromatography. See, for example, U.S. Patent No. 15 4,136,094 by Condie and Hoppe et al., Vox Sang. 25:308 (1973); Friesen et al., J. Applied Biochem. 3:164 (1981); and Walsh and O'Riordan, Irish Med. J. 75:232 (1982).

20 Condie contends that an IV administratable and stable 5 wt-% protein solution is possible without extra additives beyond the standard glycine-saline buffer. It should be noted, however, that a key step in this particular process is the pretreatment of the plasma source with colloidal silica. The present 25 process, on the other hand, does not require exposure by workers to the potential health hazard of working with silica fumes while preparing a stabilized protein solution. The other three references of the foregoing grouping rely exclusively on the 30 ion-exchange chromatography process to produce a final product said to be safe for IV administration. However, all three require that the final product, which contains very low protein concentration (1-4 wt-%), be lyophilized to meet adequate shelf 35 stability requirements. The present invention, on

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the other hand, provides a relatively low cost procedure for amply stabilizing low protein gamma globulin solutions that avoids the inconvenience and expense of lyophilization and reconstitution prior to use.

- 5 To evaluate the stability of liquid gamma globulin products, a relatively simple test has been used historically. This test is recommended by the U.S. Bureau of Biologics and involves heating the
- 10 finished product to a temperature of 57°C. and holding it at that temperature for four hours while examining the product for visual precipitates. See Code of Federal Regulations 21, Food and Drugs, 640.101a (revised April 1978). Fernandes and
- 15 Lundblad, Vox Sang. 39:101-112 (1980) report a modification of this procedure suitable for routine evaluation of potential additives. The modified procedure comprises heating approximately 2 milliliters of the test product at 57°C. for four
- 20 hours and then evaluating the percent change in degree of opalescence as measured by recording the transmittance at 580 nm with a laboratory spectrophotometer.

- The preferred buffer for purified liquid
- 25 gamma globulin products heretobefore has been glycine-saline, pH 6.4-7.2; however, this buffer has now been found to be inadequate if the protein concentration is 5 wt-% or less.

- Another problem encountered with relatively
- 30 low protein gamma globulin products which employ only glycine-saline as the stabilizer is pH control. Historically, the higher protein concentration (i.e. about 15 wt-%) has served as the principal buffering agent for the product, not either glycine or saline.
- 35 Yet the relatively lower protein concentrations do not exhibit an adequate buffering effect.

The present invention, on the other hand, provides superior pH stabilization over the historical glycine-saline buffer for purified gamma globulin solution with a protein concentration of 5 wt-% or less.

Additionally, this invention provides a relatively low cost product which is safe when injected into patients for purposes of gamma globulin therapy. The present product is effective at much lower concentrations than the previously used, stabilized products, does not require the use of hazardous chemicals like silicon dioxide to treat the gamma globulin, and does not require chemical modifications of the gamma globulin itself.

Summary of the Invention

Immunoglobulin preparations containing a stabilizing amount of histidine and having a pH value in the slightly acidic range are contemplated by the present invention. Such preparations are substantially free from anticomplement activity and contain histidine at a concentration sufficient to inhibit aggregation of the immunoglobulin that is present.

The pH value of the present preparations is at least about 6.0 but not more than 7.0, preferably about 6.2 to about 6.6, and most preferably about 6.4.

Conductivity of the present preparations is in the range of about 2 to about 4 millisiemens (mS) at 5°C., preferably about 2.5 to about 3.5 millisiemens at 5°C., and most preferably about 2.7 millisiemens at 5°C.

While any immunoglobulin can be stabilized for storage in the foregoing manner, the present invention is particularly well suited for the stabilization of human gamma globulin or IgG, the predominant serum immunoglobulin in human plasma.

To that end, a particularly preferred stabilized, sterile gamma globulin composition, exhibiting substantially no anticomplement activity, comprises an aqueous solution containing a

5 pharmacologically effective concentration of gamma globulin, histidine in a concentration sufficient to inhibit gamma globulin aggregation, and glycine in a concentration of about 0.05M to about 0.5M.

In the foregoing composition the gamma

10 globulin concentration is about 5 weight percent or less, more preferably about 0.05 to about 5 weight percent, and most preferably about 1 to about 2 weight percent.

Stabilized immunoglobulins can be

15 administered to a patient intravenously or intramuscularly and are useful for elevating the circulating antibody levels of a patient.

Detailed Description of Preferred Embodiment

Buffering ability and pH are important

20 aspects of the present invention because the solubility of most globular proteins, such as the immunoglobulins, is influenced by the pH of the system in which the protein is present. Usually the pH at which a protein is least soluble is its

25 isoelectric point, i.e., that pH value at which the protein molecule has no set electric field. At the isoelectric pH there is no electrostatic repulsion between neighboring protein molecules, and they tend to coalesce and precipitate. In practicing the

30 present invention it is believed that the immunoglobulins present, e.g., the gamma globulins, are stabilized against aggregation by maintaining the solution pH at a value other than the isoelectric pH value of the immunoglobulin molecules present.

35 To that end the amino acid histidine is utilized as a stabilizing agent. Preferably,

histidine is used together with glycine. The histidine and glycine used to stabilize the present aqueous solutions are described in detail in, for example, The Merk Index, Tenth Edition, Merk & Co., Inc. Rahway, N.J. (1983) at pages 4621 and 4359, respectively. L-Histidine is the particularly preferred stereoisomer of histidine for present purposes, but D-histidine or a racemic mixture of histidine stereoisomers are also suitable. In addition, the pharmaceutically acceptable salts of histidine and glycine can be used as well. L-histidine hydrochloride monohydrate and glycine hydrochloride are particularly preferred salts for present purposes.

The phrase "pharmaceutically acceptable salts", as used herein, refers to non-toxic alkali metal, alkaline earth metal and ammonium salts used in the pharmaceutical industry, including the sodium, potassium, lithium, calcium, magnesium and ammonium salts and the like that are prepared by methods well-known in the art. This phrase also includes non-toxic acid addition salts that are generally prepared by reacting the compounds of this invention with a suitable organic or inorganic acid. Representative salts include the hydrochloride, hydrobromide, sulfate, bisulfate, acetate, oxalate, valerate, oleate, laurate, borate, benzoate, lactate, phosphate, tosylate, citrate, maleate, fumarate, succinate, tartrate, and the like.

The concentrations of histidine and glycine in the present preparations can vary depending on such factors as salt concentration, etc. The concentration of histidine preferably is in the range of about 0.025M to about 0.2M and the concentration of glycine, if present, can be in the range of about

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0.05M to about 0.5M. In a particularly preferred composition the concentrations of histidine and glycine are about 0.05M and about 0.1M, respectively.

5 One of the advantages of the present invention is its ability to buffer the aqueous solution adequately against undesirable changes in hydrogen ion concentration (pH). The overall range of pH values for the present preparations is at least
10 about 6.0 to no greater than 7.0. In preferred embodiments, the pH value of the aqueous preparations embodying the present invention is about 6.2 to about 6.6, more preferably about 6.4.

 The buffering capacity in the present
15 preparations is believed to be provided for the most part by histidine, the only amino acid with significant buffering capacity in the desired pH range.

 Conductivity of the aqueous immunoglobulin solutions is also important for optimum stability.
20 To this end, the present solutions exhibit a conductivity of about 2 to about 4 millisiemens at 5°C., plus or minus 10%. Preferably, the solution conductivity is about 2.5 to about 3.5 millisiemens at 5°C., most preferably the conductivity is about
25 2.7 millisiemens at 5°C.

 When L-histidine hydrochloride monohydrate is used as the stabilizing agent and the final pH adjustment is made using an aqueous sodium hydroxide solution, a pH adjustment to about 6.4 will result in
30 a sodium chloride concentration in the final solution of about 0.05M and the desired conductivity of 2.7 ± 0.27 millisiemens at 5°C.

 The immunoglobulin, e.g., gamma globulin, is present in the preparations embodying this invention
35 in a pharmacologically effective concentration. The

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specific amount can vary, depending upon the intended route of administration and therapeutic use; however, in general the immunoglobulin concentration in the present preparations is about 0.05 to about 5 weight percent, preferably about 1 to about 2 weight percent.

In the case of gamma globulin, the native protein preferably is at least about 95 percent pure, more preferably about 99.5 percent pure, and has not undergone any chemical or enzymatic modification.

Gamma globulin stabilized in accordance with the present invention can be administered intravenously to patients for therapeutic purposes in single or multiple unit doses ranging from about 20 milligrams/kg/day to about 200 milligrams/kg/day over a desired time period. For intramuscular administration the individual doses are somewhat larger. The present preparations can be administered to a patient in conjunction with surgical or chemical treatments as well.

The term "unit dose" refers to physically discrete units suitable as unitary dosages for warm blooded animals, each unit containing a predetermined quantity of active material calculated to produce the desired therapeutic effect in association with the preparation of this invention. The specifications for the novel unit dose of this invention are dictated by and are directly dependent on (a) the unique characteristics of the active material, (b) the particular pharmacological or therapeutic effect to be achieved, and (c) the limitations in the art of compounding such active material for use in animals as disclosed in detail in the present specification, these being features of the present invention.

The immunoglobulins suitable for compounding the present compositions can be derived from any

source as long as the desired purity standards are met. Various immunoglobulin recovery and purification techniques are known in the art, such as ion exchange resin techniques, affinity chromatography, and the like, and are well suited for the purposes of this invention. See, for example, U.S. Patent No. 4,434,093 to Zolton et al., Applicants' copending European Patent application No. 85308922.5, and Standworth, Nature 188:156-157 (1960).

In general, the preparations of this invention are compounded by providing an aqueous solution of the desired immunoglobulin, e.g., human gamma globulin, at the requisite concentration and purity, and then combining therewith histidine in an amount sufficient to inhibit aggregation of the immunoglobulin that is present. The hydrogen ion concentration of the resulting solution is adjusted to a pH value of at least about 6.0 but not more than 7.0. Optionally, glycine can also be added in the amounts stated hereinabove. Likewise, if needed, conductivity of the resulting solution is adjusted to the hereinabove-stated limits in a manner known in the art.

The present invention is further illustrated by the following detailed examples.

EXAMPLE 1: Preparation of Human Plasma Pool

Blood units were collected from a large number of Rh-negative donors who had been previously hyperimmunized with Rh-positive red cells. Each unit was centrifuged by standard procedures to produce platelet-poor plasma. The latter was frozen individually within 4 hours of the collection. These units were stored frozen until needed. The frozen

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units were thawed and pooled together on the day the fractionation was to start. Cryo-precipitate observed to be present in the thawed plasma was removed from the plasma after pooling and by standard procedures. The obtained supernate from this process contained the desired gamma globulin and was held at 2-8°C. to await further fractionation.

EXAMPLE 2: Sample Preparation Prior to
Ion-Exchange Chromatography

The supernate (5.8 l) prepared in Example 1, above, contained a starting anti-D activity of 73.7 ug/ml and was dialyzed using a Millipore ultrafiltration unit and the column equilibration and running buffer, i.e., 0.05M imidazole-0.023M sodium chloride buffer, pH 7.5 \pm 0.1. Two volumes were exchanged. This step insured that when the plasma sample was subsequently applied to a column containing an ion-exchange resin the conductivity state of the plasma was such that the resin neither expanded nor contracted. The dialyzed sample was held at 15-20°C. prior to start of the next step. The volume of the dialyzed sample was 5.4 liters.

EXAMPLE 3: Ion Exchange Chromatography

QAE-Sephadex, A-50 anionic resin (800 g dry weight; Lot 12773), was washed, swollen and equilibrated with the buffer characterized in Example 2, above, per instruction of its manufacturer, Pharmacia Fine Chemicals, Piscataway, N.J. An equilibrated resin slurry was obtained and was loaded into a plastic column (15.0 cm high x 37.0 cm diameter) to provide a final bed volume of 16.0 liters. The dialyzed plasma sample from Example 2, above, was applied to this packed column and allowed

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to flow down through the resin bed therein at a flow rate of 1.5 liters/hour and at a temperature of $20 \pm 5^\circ\text{C}$. During this procedure the majority of the plasma proteins bind to the positively charged ion while most of the gamma globulin product does not bind and hence passes directly out the bottom of the column where it is recovered.

After all of the plasma sample had entered the resin bed, the aforesaid column buffer was continuously passed through the column. The effluent from the column was monitored for the presence of protein by measuring $\text{OD}_{280\text{nm}}$ (optical density at 280 nanometer wavelength) readings on a standard laboratory spectrophotometer. The latter readings were referenced to that of the column buffer by itself. Collections of effluent aliquots were initiated when a rising 280nm reading was observed. The collection was continued until the $\text{OD}_{280\text{nm}}$ readings returned to 20-30% of the initial baseline reading. The collected product, rich in gamma globulin, was observed to have a volume of 17.85 liters and a protein concentration of less than 0.1% by weight. The recovery of the anti-D activity was observed to be approximately 80%. The obtained dilute gamma globulin product was held at $2-8^\circ\text{C}$. until it could be concentrated to a desired antibody potency of 250 ± 25 ug Anti-D/ml.

EXAMPLE 4: Concentration of Product

The dilute product obtained in Example 3, above, was concentrated approximately 20 times using a Millipore ultrafiltration unit equipped with a 10,000-molecular weight cut-off membrane.

EXAMPLE 5: Dialysis of Final Product
With Stabilizer Solution

After concentration as described in Example 4, above, and while still in the same ultrafiltration unit, the concentrated product volume was exchanged four times with the present stabilizer solution, i.e., 0.05M histidine-0.1M glycine, pH 6.4 ± 0.1 . The final volume of the concentrated product was measured to be 1.35 liters. The protein concentration in the concentrated product was observed to be 1.4% by weight, and having a pH of 6.4 ± 0.1 and conductivity of 2.7 millisiemens ± 0.27 at 5°C.

EXAMPLE 6: Sterile Filtration and Vialing

The concentrated product stabilized in Example 5, above, and having antibody potency as reported in Example 3, above, was sterile-filtered through a 0.22 micron filter and vialled at a final volume of 1.2 ml/vial. The filled vials were stoppered, sealed, and stored at 2-8°C. for three years. Some aliquots of the concentrated and stabilized product were lyophilized prior to storage and then stored in lyophilized state. Upon repeat testing over three years the histidine-stabilized and stored product has been observed to be free of aggregates and anti complementary activity, and therefore is safe for IV injections.

Histidine and glycine are readily available in pure form and have good stability in aqueous solutions. Physiologically, they are safe and are used as dietary supplements in powdered protein preparations. However, the present invention contemplates a novel combination of histidine and glycine and, given the unpredictability of biological systems, their use as stabilizing solutes for the

purposes of the present invention was examined in safety studies as reported hereinbelow.

EXAMPLE 7: Safety Study of Histidine and

5 Glycine Solutions in Mice

A histidine and glycine solution (0.05M L-histidine hydrochloride monohydrate, 0.010M glycine, 0.003% thimerosal, in pyrogen free water; pH 6.4 \pm 0.1; conductivity 3.0 \pm 0.3mS at 0° to 5°C.)
10 was tested intramuscularly in comparison with a glycine and saline solution (0.20M glycine, 0.05M sodium chloride, 0.01% thimerosal, in pyrogen free water; pH 7.45 \pm 0.15; conductivity 3.0 \pm 0.1mS at 0° to 5°C.) and a saline control solution (0.90% sodium
15 chloride, 0.003% thimerosal, in pyrogen free water; pH 6.4 \pm 0.1).

Each solution was administered by injecting 0.2ml into the thigh muscle of mice 3 times per week for two weeks. This dosage level was calculated to
20 exaggerate the human single dose exposure by at least 1000 times. All mice survived the 2-week test and had no untoward chemical effects associated with the treatment. One mouse receiving the glycine-saline solution lost weight (one gram) during the test
25 period. This was deemed to be an insignificant weight loss.

At necropsy little reaction was observed in the intramuscular injection sites and no visceral tissue change associated with treatment was present.
30 Microscopic examination of the selected tissues (e.g., lungs, heart, liver, spleen, kidneys and injection sites from both rear legs) revealed no effect associated with the various injected solutions. The only injection site changes were
35 associated with needle puncture trauma.

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EXAMPLE 8: Safety Study of Histidine and
Glycine Solutions in Guinea Pig

The same solutions as were used in the study reported in Example 7, above, were utilized in a
5 14-day intradermal safety study with guinea pigs. A 5 wt-% L-histidine hydrochloride monohydrate solution in pyrogen-free water, adjusted to a pH value of 6.4 ± 0.1 , was utilized as well. Each test solution was injected into five adult female guinea pigs
10 (Hartly Descendant; 400+ grams in weight) that had been quarantined for at least 7 days.

The back of each animal was shaven five days prior to the first injection. Each animal was dosed three times (on Day 1, Day 10 and Day 14) with 0.1 cc
15 of test solution, on the back, near the midline.

The animals were observed daily for any abnormality. The injection sites for each animal were observed at 24 and 48 hours post injection for redness or swelling.

20 During the course of this study no reaction to the test solutions was noted. While one guinea pig receiving the histidine-glycine test solution became ill with a cold during the test period (diarrhea, weight loss, chills, nasal discharge),
25 this animal had no observable reaction to the injected test solution.

Various comparative studies have been carried out with the stabilized preparations embodying the present invention. These studies and
30 the results thereof are set forth in Tables I through V that follow.

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Table I

HIGH PERFORMANCE LIQUID CHROMATOGRAPHY
(HPLC) PROFILE OF STABILIZED LIQUID
LOW PROTEIN GAMMA GLOBULIN PRODUCT

5

		<u>HPLC Results (c)</u>		
		<u>% Monomer</u>	<u>% Dimer</u>	<u>% Aggregate</u>
10	Control (a)	84.242	15.009	0.112
	Purified human gamma globulin (b) stabilized by 0.05M L-histidine-	100.000	0.000	0.000
15	0.1M glycine, pH 6.4, stored as liquid product at 2-8°C.			
20	Purified human gamma globulin (b) stabilized by 0.05 L-histidine-0.1M glycine, pH 6.4, stored as a lyophilized product at 2-8°C.	100.000	0.000	0.000
25	_____			

(a) The control was a commercially available, 15.0% by weight protein human gamma globulin product derived from cold alcohol process. This sample was expected to contain measurable levels of all three components.

30

(b) Method used to produce purified gamma globulin was ion-exchange chromatography as described in Examples 1-6, above. The same source was used for both samples.

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The final protein concentration was 1.4% by weight.
Both samples were stored unopened 24 months at 2-8°C.

- (c) High performance liquid chromatography (HPLC)
5 assay was done using a TSK-3000 SW column based on the
published report by Bertolini, Vox Sang. 43:87 (1982).
The flow rate was 0.5 ml/min and the detector wavelength
was set at 280nm. The aggregates, where present, were
observed to elute at approximately 13.6 minutes followed
10 by a dimer peak at approximately 16.5 minutes and a
monomer peak at approximately 19.5 minutes.

Data in Table I show that during a two-year
storage period L-histidine stabilized gamma globulin
15 does not exhibit appreciable aggregate formation.

Table II

RESULTS FROM HEAT-STRESS TESTING^(a)

20

- I. Comparison of Various Buffers on the Stability of
Purified Human Gamma Globulin (IgG) Derived from
the Standard Cold Alcohol Process

25

<u>Sample</u>	<u>% Transmission Change</u>
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"A"	3 wt-% IgG in 0.2M glycine- 0.05M sodium chloride, pH 6.4.	> 90.0
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30

"B"	3 wt-% IgG in 0.025M histidine- 0.2M glycine, pH 6.4.	6.7
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"C"	3 wt-% IgG in 0.05M histidine- 0.2M glycine, pH 6.4.	6.7
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II. Comparison of Various Buffers on the Stability
of Purified IgG Derived from an Ion-Exchange
Chromatography System

5	<u>Sample</u>	<u>% Transmission Change</u>
	"D" 2.0 wt-% IgG in 0.2M glycine- 0.05M sodium chloride, pH 6.4.	24.0
10	"E" 2.0 wt-% IgG in 0.1M histidine, pH 6.4.	7.6
	"F" 2.0 wt-% IgG in 0.05M histidine- 0.1M glycine, pH 6.4.	5.4

15

III. Relative Effect of Various Agents on Heat-Stress
Stability of Purified IgG Derived from an
Ion-Exchange Chromatography System

20	<u>Sample</u>	<u>% Transmission Change</u>
	"G" 2.5 wt-% IgG stabilized by 0.2M glycine-0.05M sodium chloride, pH 6.4.	23.6
25	"H" 2.5 wt-% IgG stabilized by 0.2M glycine-0.05M sodium chloride and 0.5 wt-% tryptophan, pH 6.4.	23.6
30	"I" 2.5 wt-% IgG stabilized by 0.2M glycine-0.05M sodium chloride and 0.5 wt-% phenylalanine, pH 6.4.	20.8

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5	"J"	2.5 wt-% IgG stabilized by 0.2M glycine-0.05M sodium chloride and 0.001 wt-% cysteine, pH 6.4.	19.7
10	"K"	2.5 wt-% IgG stabilized by 0.2M glycine-0.05M sodium chloride and 10 wt-% maltose, pH 6.4.	5.0
15	"L"	2.5 wt-% IgG stabilized by 0.2M glycine-0.05M sodium chloride and 10 wt-% sucrose; pH 6.4.	6.5
20	"M"	2.5 wt-% IgG stabilized by 0.1M glycine-0.05M histidine, pH 6.4.	5.4
	IV.	Evaluation of the Effect of pH on Stabilizer Performance	

	Sample	Final pH	% Transmission Change
25	"N" 2.5 wt-% IgG in 0.05M histidine- 0.1M glycine	6.4	5.4
30	"O" 2.5 wt-% IgG in 0.05M histidine- 0.1M glycine (b)	7.2	69.0

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"p" 2.5 wt-% IgG in
0.02M sodium
phosphate

6.4

off-scale; noted
17.0% change after
2 hours.

5

(a) Heat stress testing was done according to the
procedure outlined by Fernandes and Lundblad in Vox
10 Sang. 39, 101 (1980).

(b) pH Adjustment was made by the addition of aqueous
NaOH solution, q.s.

15

The data set forth in the above Table
demonstrate that a glycine-saline buffer is not
adequate for the stabilization of purified liquid
gamma globulin products were the protein
concentration is relatively low. The foregoing data
20 also underscore the unexpected advantages from
utilizing histidine to stabilize such products at a
pH value in the slightly acidic range.

25

30

35

Table III

MEASUREMENT OF ANTICOMPLEMENTARY ACTIVITY
IN AGED STABILIZED LOW PROTEIN PRODUCT

5

Anticomplementary Titer^(a)

Control Sample 1:64

10 Stabilized Product^(b)
(stored 6 months at 2-8°C.) less than 1:2

Stabilized Product^(b)
(stored 2 years at 2-8°C.) less than 1:2

15

(a) The anticomplementary activity of the above-identified globulin was determined by the method described by Friesen et al., J. Applied Biochemistry, 3:164-175 (1981). Serial twofold dilutions of the IgG solution were incubated with 5 CH₅₀ units of complement. Following overnight incubation at 4°C., 1% sensitized sheep blood cells were added and incubated at 37°C. for 30 min. The anticomplementary activity was expressed as the protein concentration milligrams per milliliter of the dilution showing at least 50% hemolysis.

(b) This product was prepared with ion-exchange purified human gamma globulin as described in Examples 1-6. Final protein concentration of sample was 1.4% by weight. The product was stored in a sealed container as a liquid.

35 The data in Table III show that the compositions stabilized in accordance with the

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present invention exhibit substantially no
anticomplementary activity.

Table IV

5

EFFECT OF IgG CONCENTRATION ON BUFFERING
CAPACITY OF STANDARD GLYCINE-SALINE BUFFER

	<u>% IgG (by Weight)</u> (a)	<u>pH</u> (b)
10	0	6.40
	0.40	6.75
	1.00	6.90
	1.80	6.95
15	2.50	6.95
	2.75	7.00
	3.80	7.00

20 (a) Samples used ion-exchange purified human gamma globulin as described in Examples 1-5. Dried product was added in various amounts to the standard glycine-saline buffer, 0.2M-0.05M, pH 6.4.

25 (b) pH Was measured at 20-25°C. by standard laboratory procedures.

30 The data in Table IV demonstrate that the buffering capacity of the standard glycine-saline buffer is inadequate for the tested protein concentrations. Had the buffer capacities been adequate, no change in the pH value of the samples would have been observed.

35

Table V

EFFECT OF DIFFERENT LEVELS OF
GLYCINE IN FINAL STABILIZING BUFFER

5

	<u>Sample (a)</u>	<u>HPLC Results (b)</u>		
		<u>% Monomer</u>	<u>% Dimer</u>	<u>% Aggregate</u>
10	Purified human gamma globulin stabilized by 0.05M histidine-0.01M glycine, pH 6.4	99.820	0.170	0.004
15	Purified human gamma globulin stabilized by 0.05M histidine-0.47M glycine, pH 6.4.	99.882	0.111	0.002

20 (a) Method used to produce purified gamma globulin was ion-exchange chromatography as shown in Examples 1-6, above. The same source was used for both samples. The final protein concentration was 1.2% by weight. Both samples had been stored 11 months unopened at 2-8°C. prior to the above testing.

30 (b) High performance liquid chromatography assay was done using a TSK-3000SW column based on the published report by Bertolini, Vox Sang. 43:87 (1982). The flow rate was 0.5 ml/min and the detector wavelength was set at 280nm. The aggregates, where present, were observed to elute at approximately 13.6 minutes, followed by a dimer peak at approximately 16.5 minutes and a monomer peak at approximately 19.5 minutes.

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The data in Table V show that within the range of glycine concentrations of about 0.01M to about 0.5M the stabilizing effect of the present solutions is not dependent upon glycine concentration.

5 While the present invention has been described and exemplified primarily with reference to human gamma globulin, the stabilization techniques described hereinabove are applicable to other immunoglobulins as well. For example, whole native
10 antibodies prepared using hybridoma technology such as that described in Niman et al., Proc. Nat'l Acad. Sci. U.S.A. 80:4949-4953 (1983), and in Hybridoma Techniques, EMBO, SKMB Course 1980, Basel, Cold Spring Harbor Laboratory, Cold Spring Harbor, Maine,
15 can also be advantageously stabilized as taught hereinabove, stored and utilized, inter alia, for diagnostic purposes. Moreover, monoclonal antibodies can be obtained not only from in vitro hybridoma culture supernatants, but also in generally more
20 concentrated form from ascites fluid of mammals into which the desired hybridoma has been introduced. Again, the production of monoclonal antibodies using ascites fluid is well known in the art. However, such antibodies can be advantageously stabilized and
25 stored for later use utilizing the features of this invention.

 The foregoing discussion and the specific examples are intended as illustrative of the present invention and are not to be taken as limiting. Still
30 other variations within the spirit and scope of this invention are possible and will readily present themselves to those skilled in the art.

35

CLAIMS

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1. A stabilized immunoglobulin preparation comprising an aqueous solution of an immunoglobulin in a pharmacologically effective concentration; said solution being substantially free from anticomplement activity, containing histidine at a concentration sufficient to inhibit aggregation of said immunoglobulin, having a pH value of at least about 6.0 but no greater than 7.0; and a conductivity of about 2 to about 4 millisiemens at 5°C.
2. The preparation of claim 1 wherein the immunoglobulin concentration is no more than about 5 weight percent.
3. The preparation of claim 2, wherein the immunoglobulin concentration is about 0.05 to about 5 weight percent.
4. The preparation of claim 3, wherein the immunoglobulin concentration is about 1 to about 2 weight percent.
5. The preparation of any one of claims 1 to 4, wherein the histidine concentration is about 0.025 M to about 0.2 M.
6. The preparation of claim 5, wherein the histidine concentration is about 0.05 M.
7. The preparation of any one of claims 1 to 6, wherein the pH value is about 6.2 to about 6.6.
8. The preparation of claim 7, wherein the pH value is about 6.4.
9. The preparation of any one of claims 1 to 8, the conductivity of which is about 2.5 to 3.5 millisiemens at 5°C.
10. The preparation of claim 9, the conductivity of which is about 2.7 millisiemens at 5°C.

11. The preparation of any one of claims 1 to 10, wherein the immunoglobulin is gamma globulin.
12. The preparation of any one of claims 1 to 10, wherein
5 the immunoglobulin is monoclonal antibody.
13. The preparation of any one of claims 1 to 12, wherein the histidine is L-histidine.
- 10 14. The preparation of any one of claims 1 to 13, additionally including glycine at a concentration of about 0.05 to 0.5M.
- 15 15. The preparation of claim 14, wherein the glycine concentration is about 0.1M.
16. A method for the stabilization of an immunoglobulin which comprises the steps of
providing an aqueous solution of an immunoglobulin;
combining with said solution histidine in an amount
20 sufficient to inhibit aggregation of the immunoglobulin present;
adjusting the hydrogen ion concentration of the
resulting solution to a value of at least about 6.0 but
not more than 7.0; and
25 adjusting, if necessary, the conductivity of the resulting solution to a value of about 2 to about 4 millisiemens at 5°C.
- 30 17. The method in accordance with claim 16 wherein the immunoglobulin is human gamma globulin having a purity of at least about 95 percent, wherein the histidine is L-histidine; and wherein the pH value of the resulting solution is adjusted to about 6.4 and the conductivity of the resulting solution is adjusted to about 2.7
35 millisiemens at 5°C.
18. The method in accordance with claim 16 or claim 17 wherein additionally glycine is combined with said solution in an amount sufficient to provide in the resulting solution a glycine concentration of about 0.05M to about 0.5M.

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19. The method in accordance with claim 16 wherein the immunoglobulin is human gamma globulin having a purity of at least about 99.5 percent and present at a concentration of about 0.05 to about 5 weight percent, wherein L-histidine is combined with said solution in an amount providing a solution concentration of about 0.025M to about 0.2M, wherein additionally glycine is combined with said solution in an amount providing a solution concentration of about 0.05M to about 0.5M, wherein pH of the resulting solution is adjusted to a value of about 6.4, and wherein conductivity of the resulting solution is adjusted to 2.7 millisiemens at 5°C.
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